



Impact of water activity on the effectiveness of high solids enzymatic hydrolysis of lignocellulose

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Enzymatic hydrolysis at high solids conc

- VHG fermentation is today's standard in the fuel ethanol industry = 12-15% ethanol is common
- For lignocellulosic feedstocks this would require 40+ % initial solids
- Technical challenging to operate processes at solids concentrations above 15% DM biomass e.g. mixing problems during hydrolysis
- High solids concentration is important for process economics:
 - Higher sugar and ethanol concentrations mean less energy for distillation
 - Reduced water usage and less water for waste treatment
 - Reduced CAPEX, higher plant productivity



The barrier broken – free fall mixing



Jørgensen *et al.*, (2007) Liquefaction of lignocellulose at high solids-concentrations. *Biotechnol. Bioeng.* 96(5):862-870.



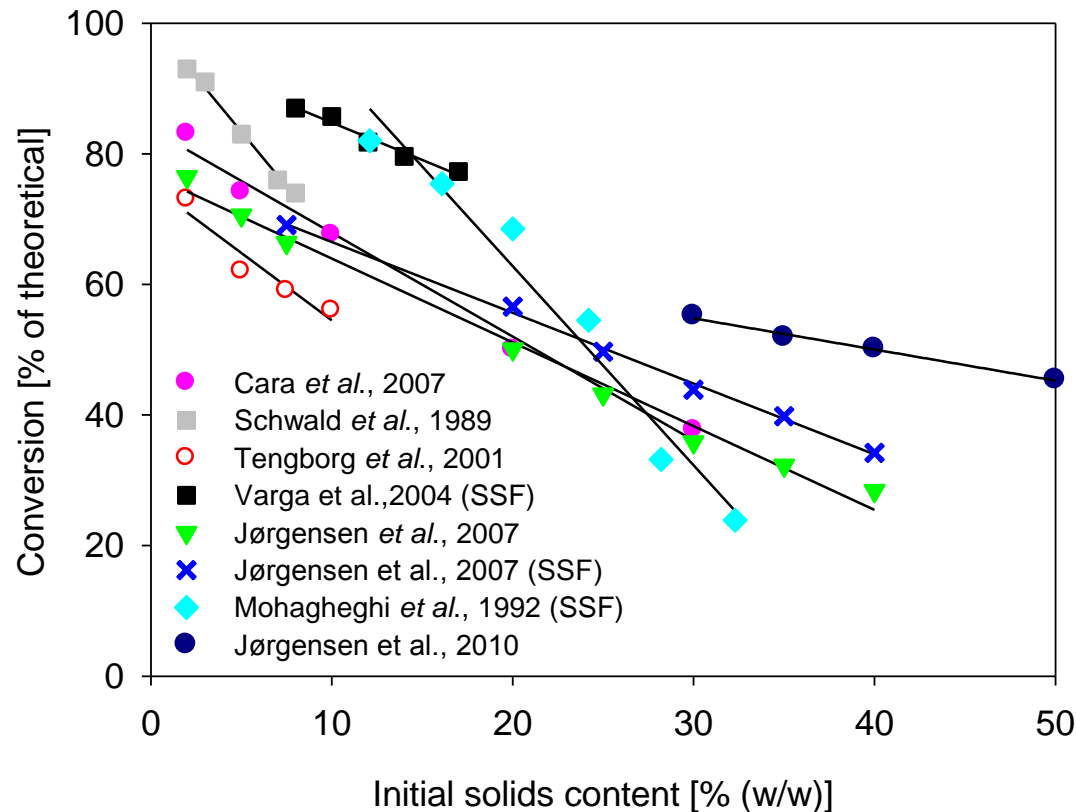
Pretreated wheat straw 30 % DM
Final ethanol concentrations up to 8 %
(w/w) or 10% (v/w)



Upscaling and proof of technology



It works BUT it could be better!



Decreasing conversion at increasing initial solids content

Observed on various feedstocks in hydrolysis but also SSF



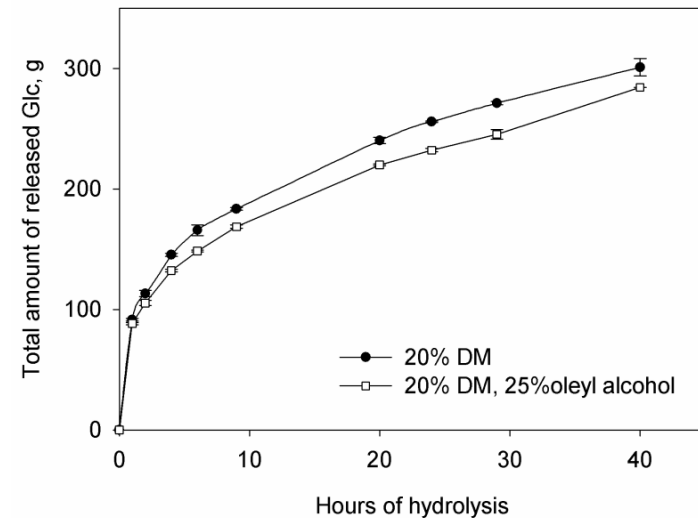
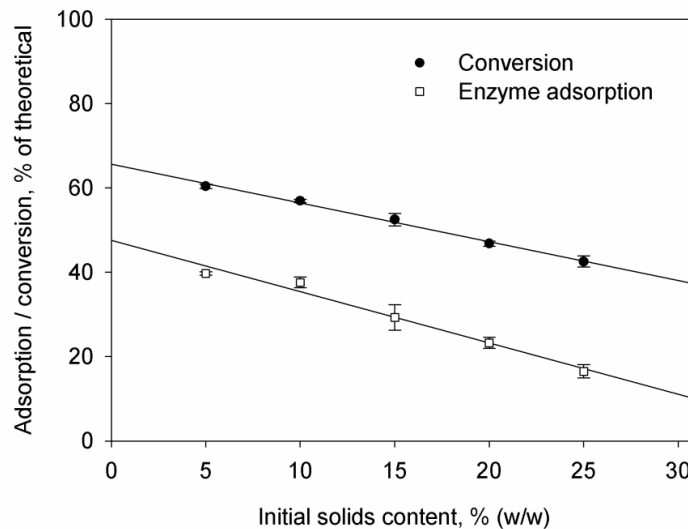
The solids effect

Factors limiting conversion:

- Product inhibition by glucose and cellobiose already during early stage of hydrolysis/liquefaction
- Inhibition by other sugars (classical inhibition)
- Inhibition by ethanol in SSF
- Stress of yeast due to high initial concentration of sugar and inhibitors originating from pretreatment
- Mass transfer limitations
- Effect of reduced (free) water



Initial study of factors that limits conversion



- Product inhibition does play a significant role
- The increasing concentration of sugars seems to affect enzyme adsorption
- Partial substituting water with oleyl alcohol revealed that water does play an role during hydrolysis – both as substrate and as solvent

Kristensen *et al.*, (2009) Yield-determining factors in high-solids enzymatic hydrolysis of lignocellulose. *Biotechnol. Biofuels* 2:1, 11.



Water activity

- Water activity is a measure of energy state of the water in a system
- Activity is relative to that of pure water
 - $a_w = p(aq)/p^*(l)$
 - Pure water has activity 1
 - Addition of a solute to water (l) always lowers its thermodynamic activity
- Water activity much used in the food industry as a indicator of stability of food towards microbial growth – low a_w depresses growth
- Factors that impact (depress) a_w in a system:
 - Colligative effects of dissolved species (e.g. salt or sugar) - dipole-dipole, ionic, and hydrogen bonds
 - Capillary effects - changes in the hydrogen bonding between water molecules
 - Surface interactions with chemical groups on undissolved ingredients - dipole-dipole forces, ionic bonds , van der Waals forces (hydrophobic bonds), and hydrogen bonds



LF-TD-NMR to study the state of water

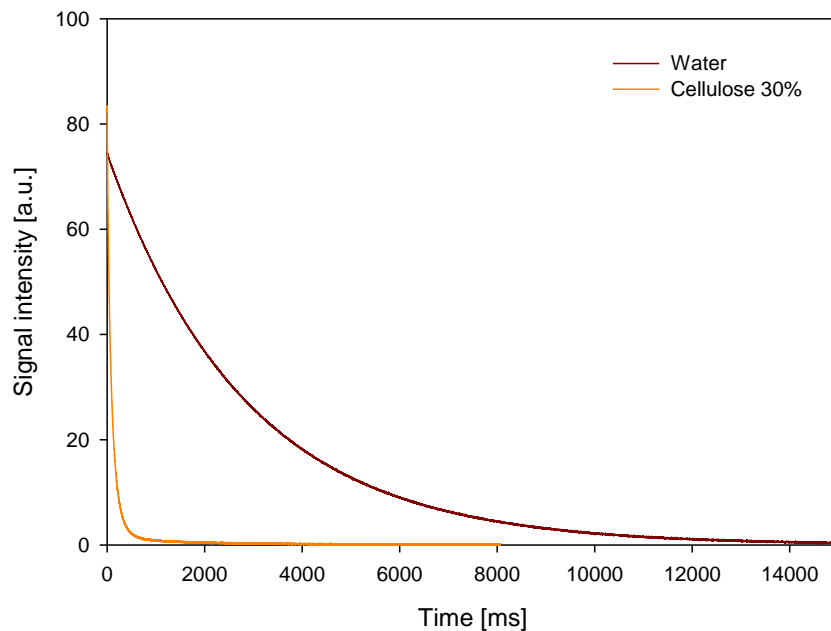
- Low Field Time Domain NMR
- Probes the states of water in the sample by measuring proton relaxation
- All protons observed at the same resonance frequency (20 MHz) and discriminated by variations in relaxation time
- The T_2 (spin-spin) relaxation time of the hydrogen nuclei depends on how free it is to move, which is determined by the environment
- Longer relaxation times means more free water
- States: tightly bound, loosely bound, and free

Felby *et al.*, (2008) Cellulose–water interactions during enzymatic hydrolysis as studied by time domain NMR. *Cellulose* 15:703-710.

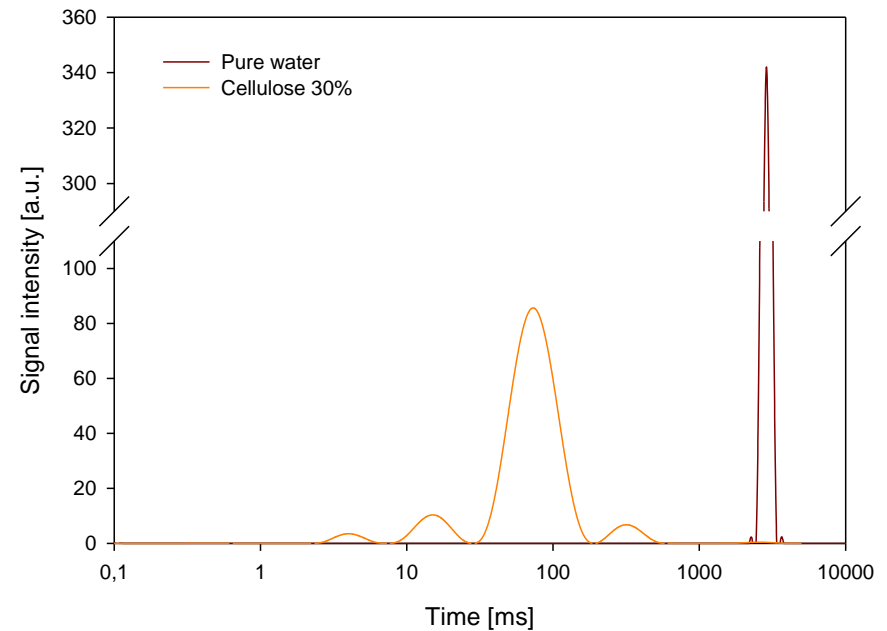


LF-TD-NMR of water and cellulose

T₂ Relaxation curve (CPMG)



T₂ distribution (CONTIN)

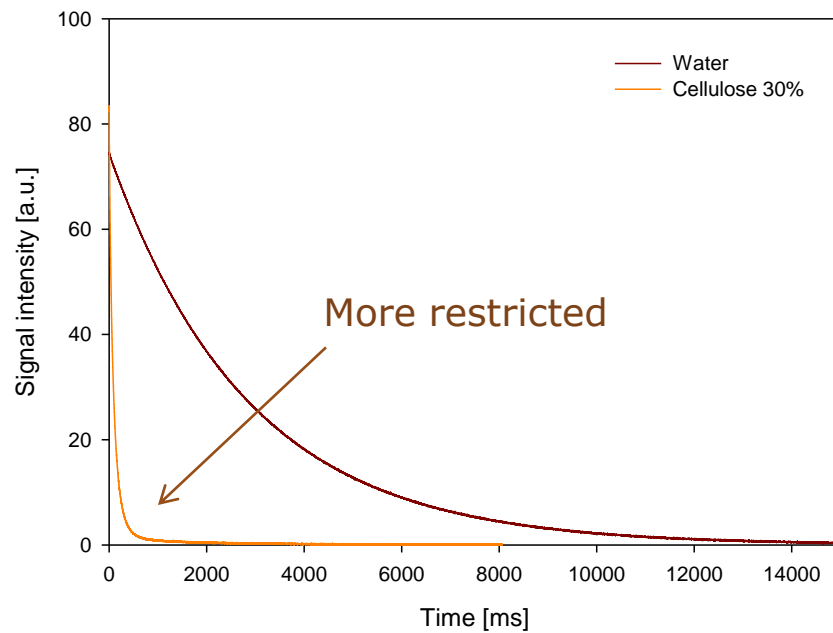


NMR of pure water and a 30% slurry of water and cellulose
Measure transverse (spin-spin) relaxation time (T₂) using the
Carr-Purcell-Meiboom-Gill (CPMG) sequence and CONTIN T₂
distribution profiles

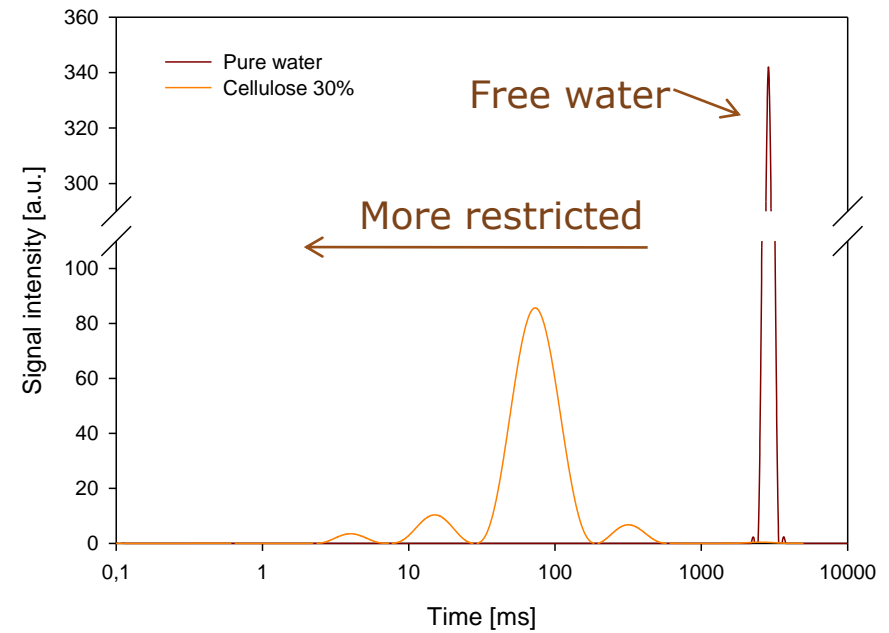


States of water

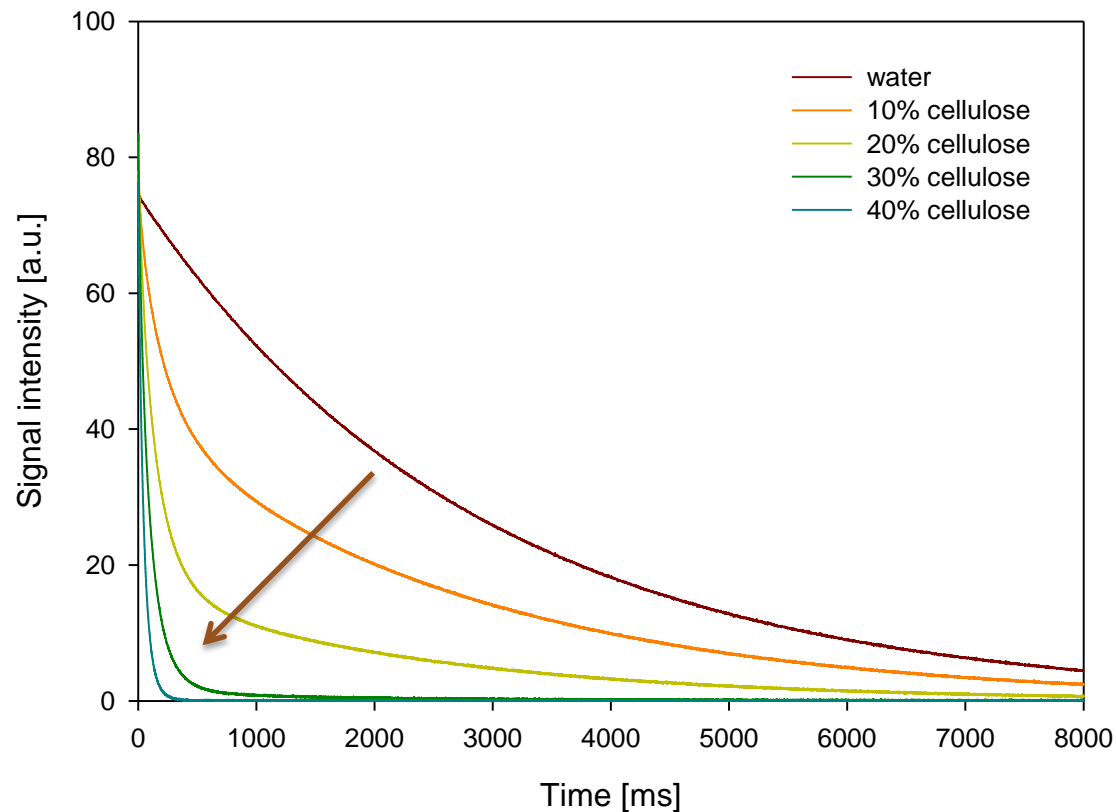
T₂ Relaxation curve (CPMG)



T₂ distribution (CONTIN)



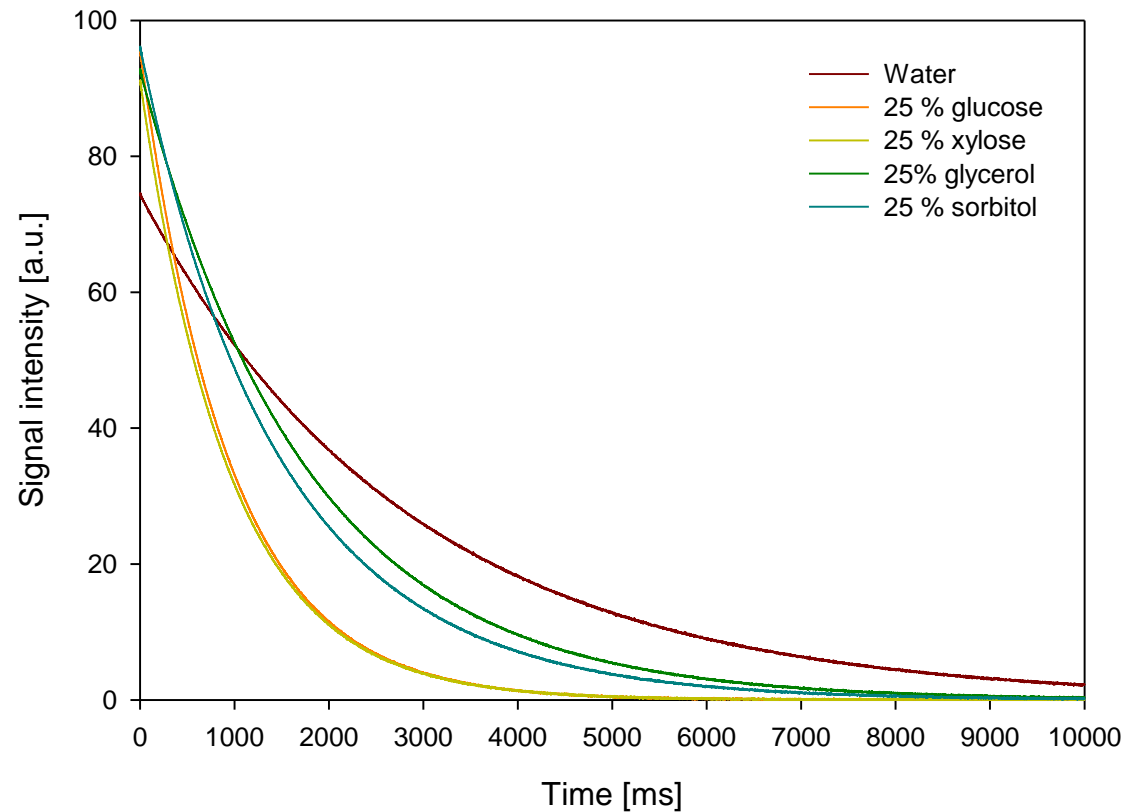
Cellulose-water system



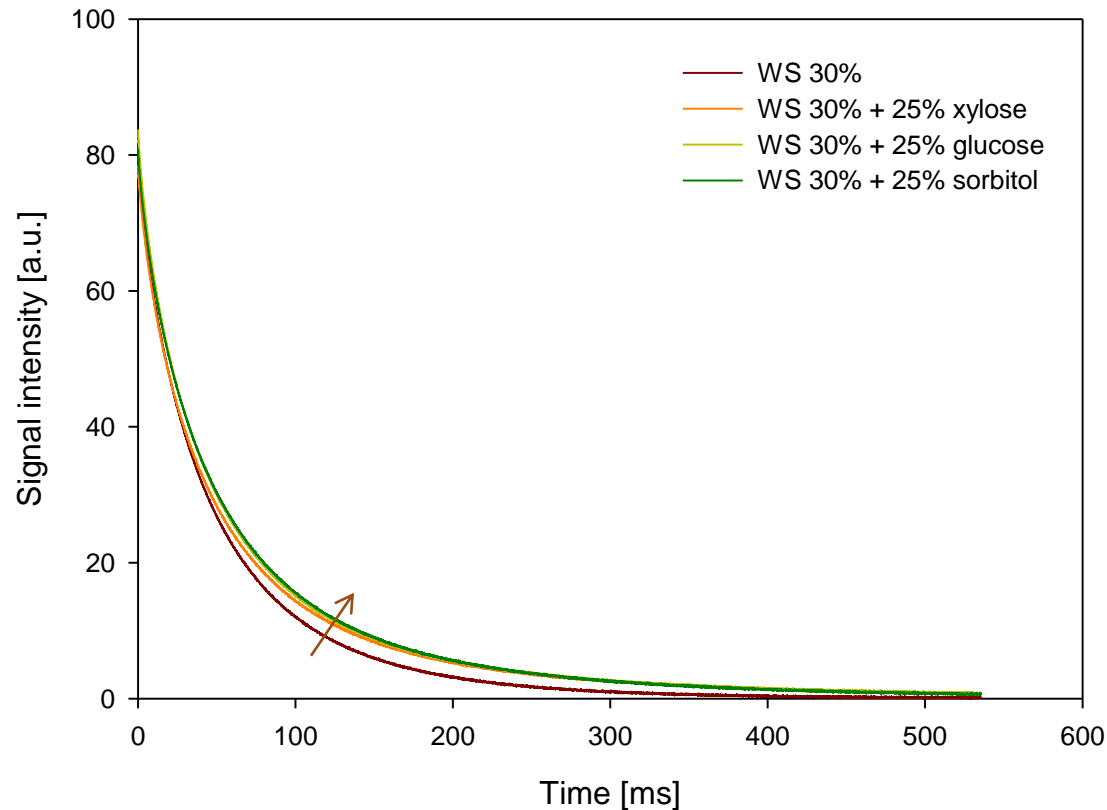
Water more restricted as the solids content increase



Sugars-water system



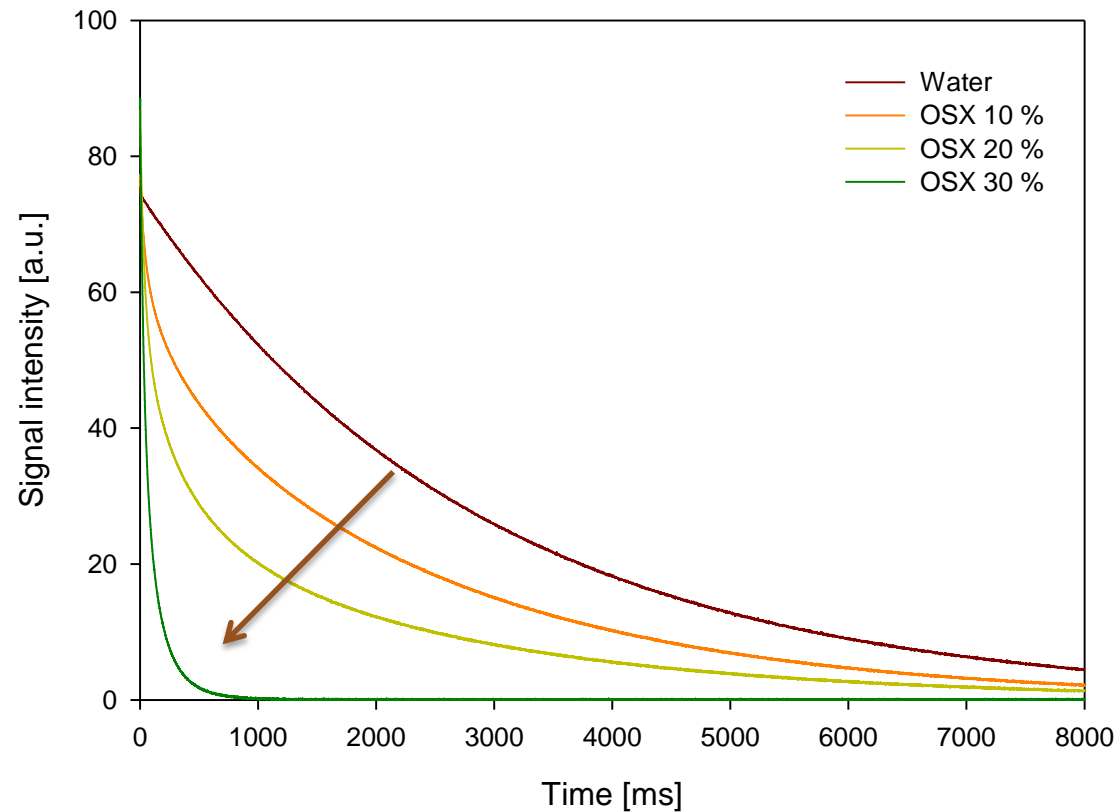
Effect of soluble sugars in WS system



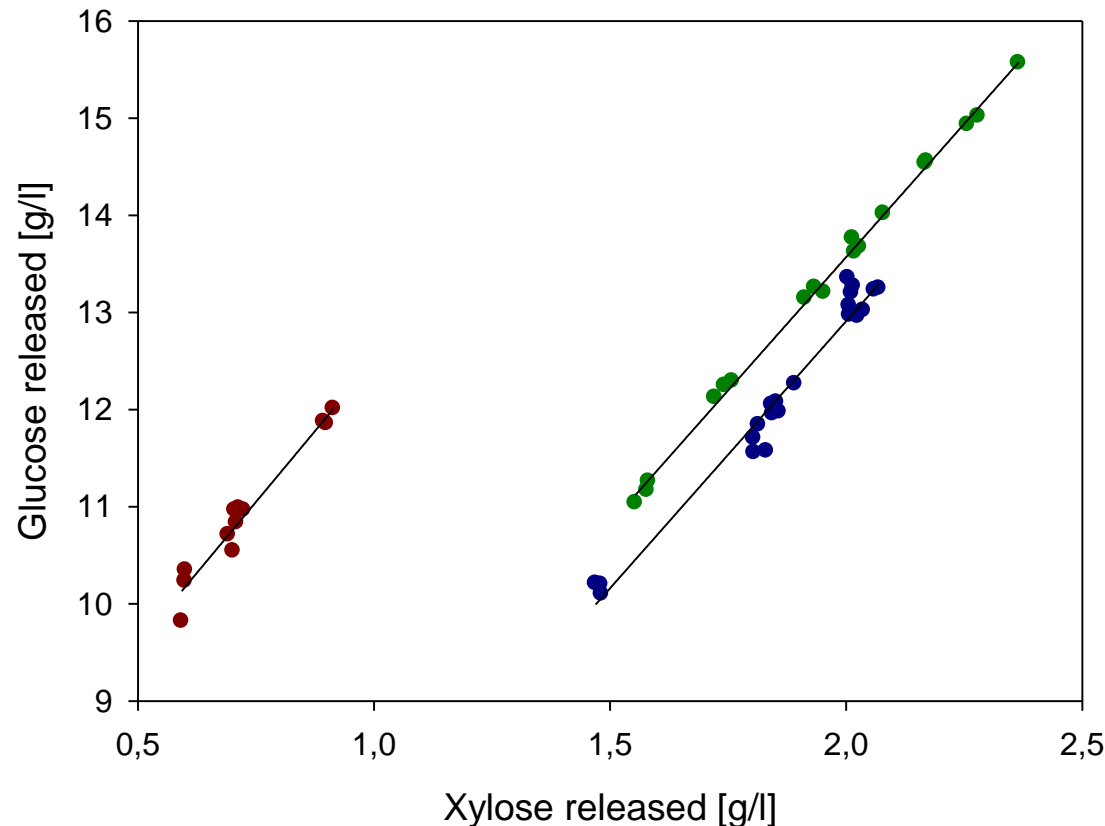
Addition of soluble sugars to system of wheat straw results in some water pools getting less constrained



Xylan-water system



Correlation between cellulose and xylan conversion



Inbicon pretreated wheat straw

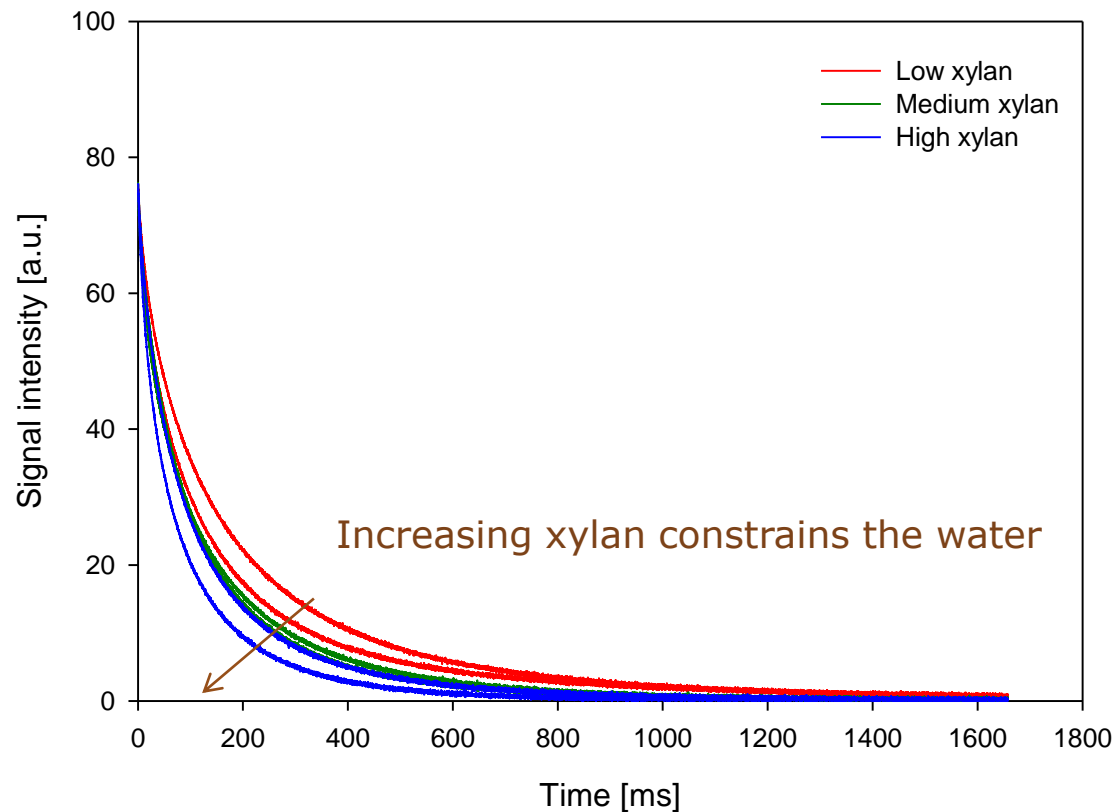
Red – low xylan, different xylanase preparations

Green – medium xylan, increasing loading of a xylanase preparation

Blue – medium xylan, different combinations of xylanase and acetyl xylan esterase



Effect of xylan in pretreated biomass



Pretreated wheat straw from Inbicon

Low xylan – 4.0-4.1%

Medium xylan – 7.9-8.2%

High xylan – 13.1-13.4%



Observations

- Adding any solute lowers a_w and NMR data show that water gets more constrained
- There is a correlation between decreased a_w and lower conversion rates
- Insoluble solids such as cellulose do not lower a_w but NMR data do show more constrained water
- Distribution of water pools depend on amount of added solute and which type
- Some sugars/sugar polymers seem to restrict water, e.g. xylan, more than cellulose



Hypothesis

- When water gets more constrained the availability of water at the cellulose surface is altered
- At low water to substrate levels water solubles may (may) pull substrate bound water away from surface of cellulose
- Surface water changes may effect enzyme (cellulase) functionality e.g. the binding of enzyme to the surface
- Previous interpretation of sugars, sugar alcohols and ethanol to be inhibitors may be explained by their ability to bind water and indirectly reduce the enzyme performance



Conclusion

- The function of the cellulase system is affected by the state of water in the system
- At high solids concentration the presence of solutes (soluble sugars, sugar polymers, sugar alcohols and ethanol) can depress the conversion rate by constraining the water and pulling surface bound water away from cellulose
- Water is definitely important for the enzyme performance (especially for enzymes working on a insoluble substrate)
- LF-TD NMR promising technique that combined with other (traditional) analysis methods can expand our understanding of how the enzyme system is working





Thank you for your attention

Acknowledgement

